

# SELEX and Artificial Ribozymes

## Some definitions

**SELEX:** Systematic Evolution of Ligands by EXponential Enrichment (alternatively: in vitro selection, in vitro evolution)

**Aptamer:** nucleic acid ligand (from Aptus: to fit)

*Jack Szostak (Boston, MA)*

*Jerry Joyce (La Jolla, CA)*

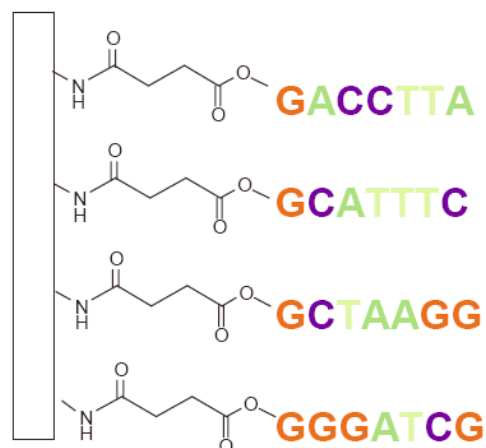
*Larry Gold (Boulder, CO)*

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## Combinatorial RNA libraries

- Straight-forward synthesis: use of nucleotide mixtures in automated solid phase synthesis,
- Enormous complexities: typically  $10^{14}$  to  $10^{16}$  different molecules,
- Nucleic acids can be enzymatically copied.

# Library synthesis - basics



Combinatorial Chemistry!

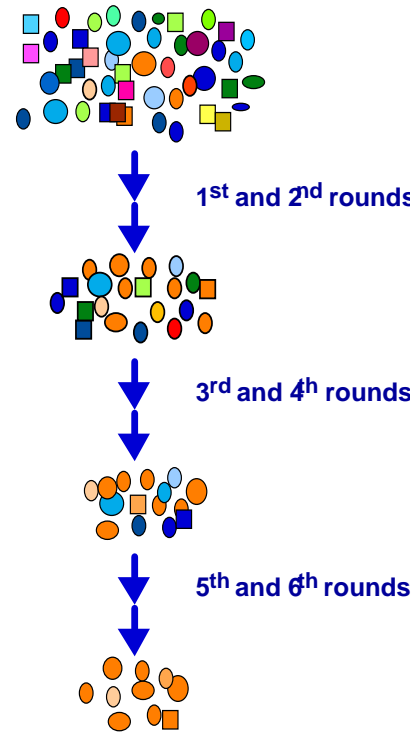
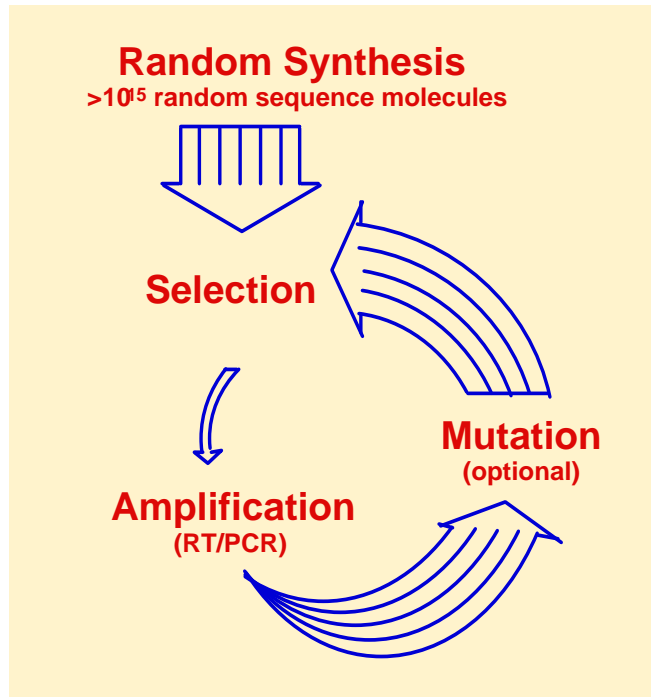
**To meditate!**

Size of the RNA fragment (nt)	Number of possible sequences	Mass of RNA required to explore all the possible sequences
20	$1.1 \times 10^{12}$	0.012 $\mu\text{g}$
<b>30</b>	<b><math>1.5 \times 10^{18}</math></b>	<b>0.018 g</b>
40	$1.2 \times 10^{24}$	25 kg
100	$1.6 \times 10^{60}$	$8.4 \times 10^{37}$ kg

1 mole =  $6.022 \times 10^{23}$  molecules

Mass of the solar system:  $2 \cdot 10^{30}$  kg

# In vitro selection and evolution techniques



*Ellington & Szostak (1990), Tuerk & Gold (1990), Beaudry & Joyce (1992)*

## SELEX

### (I) Criteria of selection

- (1) Purification by affinity chromatography (ligand, protein etc.)
- (2) Purification by gel shift (protein, RNA ligand, peptide etc.)
- (3) RNA self-modification leading to a change of size or attachment of biotin (followed by gel purification or streptavidin affinity column etc.)

### (II) Amplification

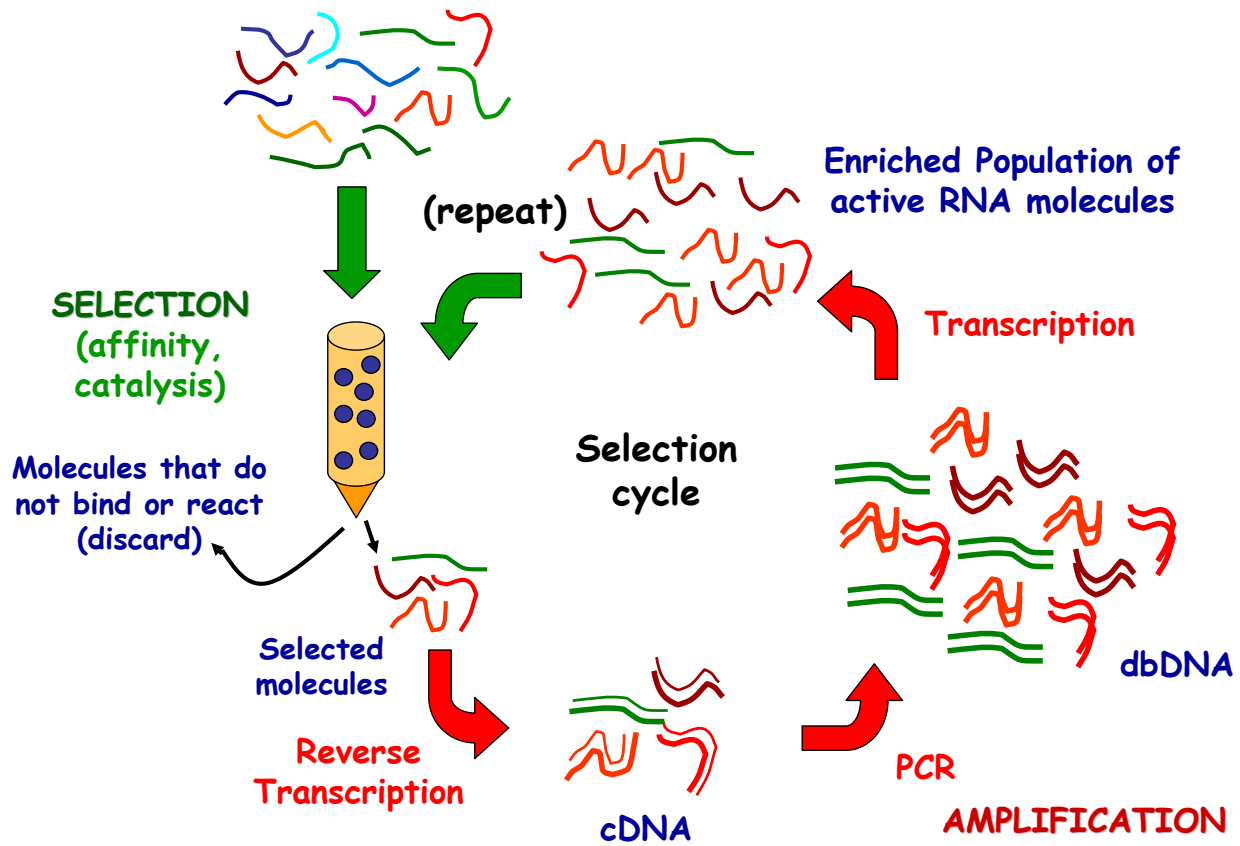
Once the selection is performed, the molecules that have been selected are amplified (first step: reverse transcription; second step: PCR)

### (III) Mutagenesis

To improve sub-optimal population of molecules (evolution) by introducing random mutations

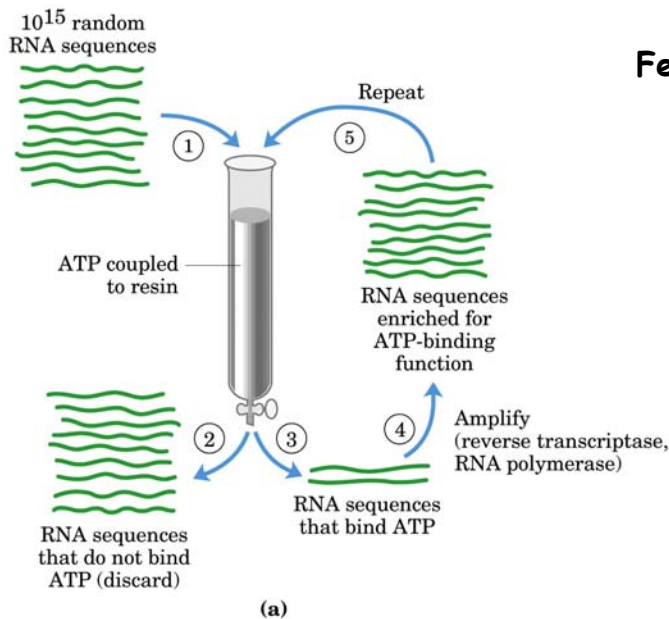
(same length, different sequence)  
**RNA population**  
 ( $10^{15}$  sequences)

# The SELEX method



## In vitro selection and evolution methods (SELEX)

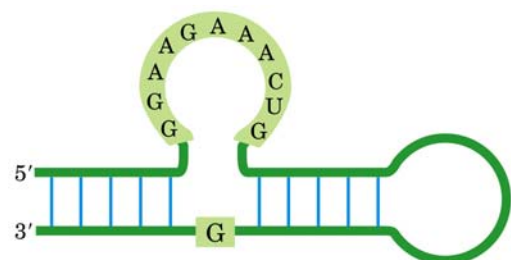
*Jack Szostak (Boston, MA)  
 Jerry Joyce (La Jolla, CA)  
 Larry Gold (Boulder, CO)*



### Few definitions

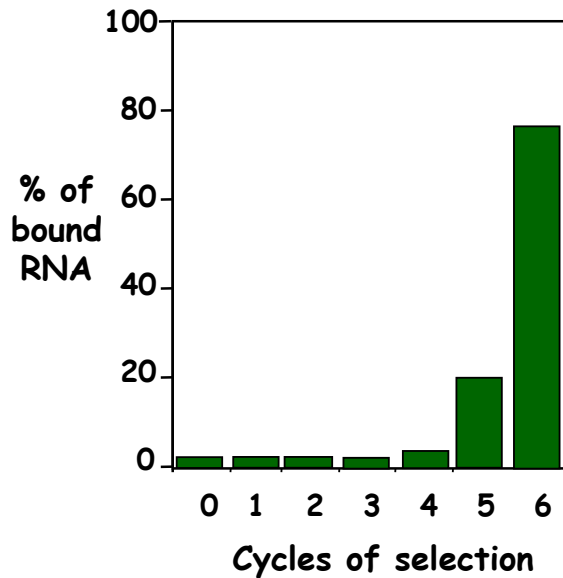
**SELEX:** Systematic Evolution of Ligands by Exponential Enrichment

**Aptamer:** nucleic acid ligand (from Aptus: adjust)



**Example: Selection of an ATP aptamer**  
 (Sassanfar & Szostak (1993) Nature 364, 550)

Example of enrichment in active molecules of a population of RNA (eg bound to a dye column) by SELEX



### little exercise

In theory, if the background noise is **1%**, then the **enrichment factor** in active molecules (with an affinity of 100%) is **100 fold at each cycle.**

If the starting population has  **$10^{13}$  different** molecules and if a signal of **20%** appears at the **5<sup>th</sup> cycle**, then there are more than **200 active molecules** (with an affinity of 100%) in the population.

$$0,2 \times 10^{13} / 100^5 = 200$$

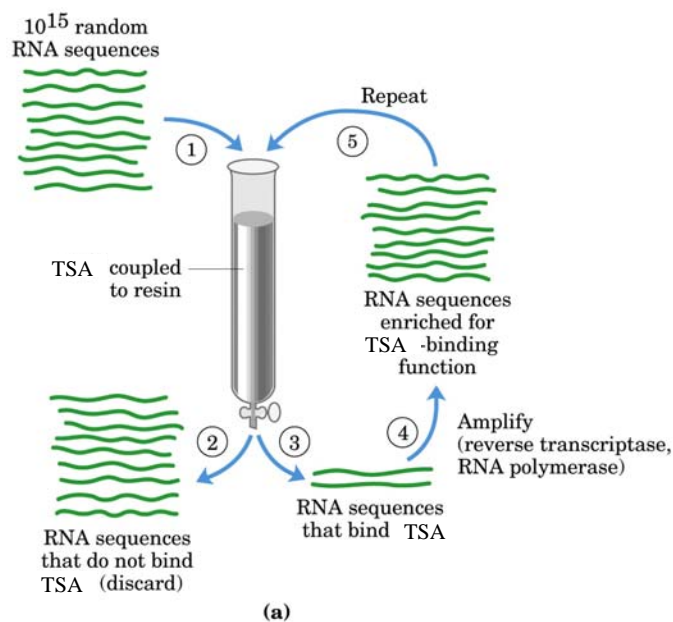
## Principles of library design



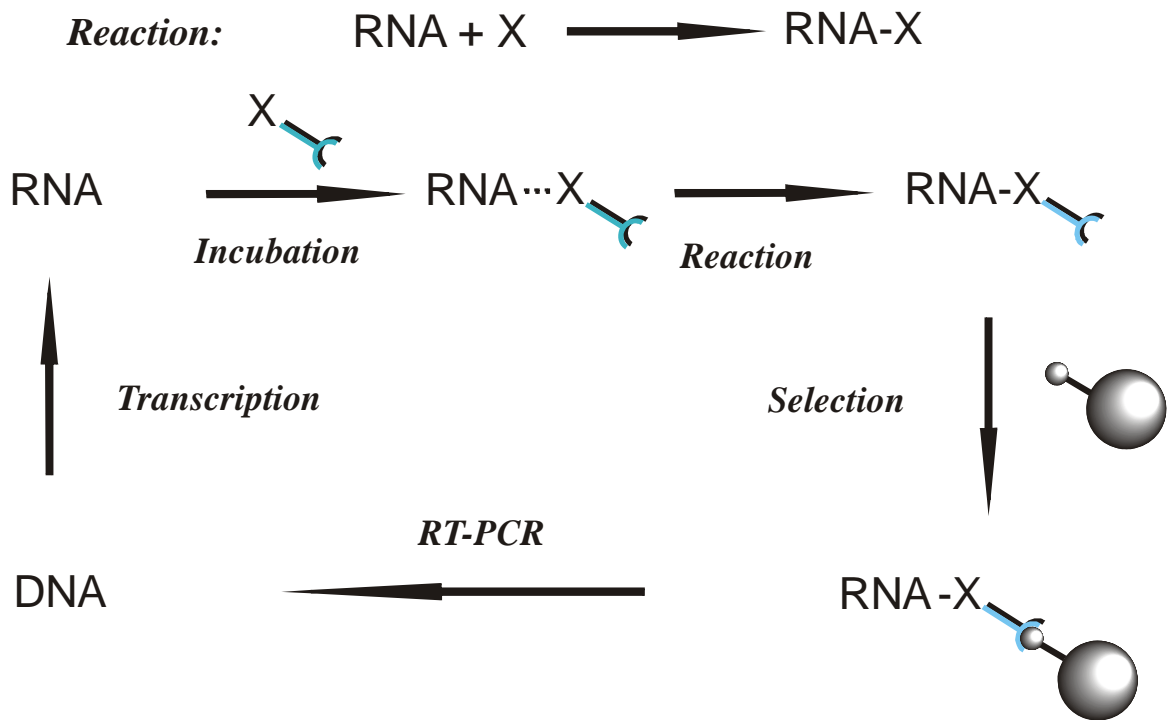
# Artificial ribozymes - methodology

- SELEX against transition state analogs
- Direct selection
- Direct selection with tethered reactants

## Selection against transition state analogs

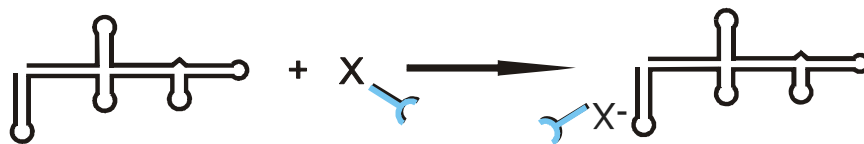


# Direct selection of RNA catalysts



# Direct selection of RNA catalysts

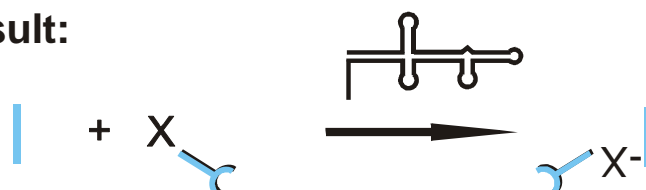
Result of the selection:



Engineering



Result:



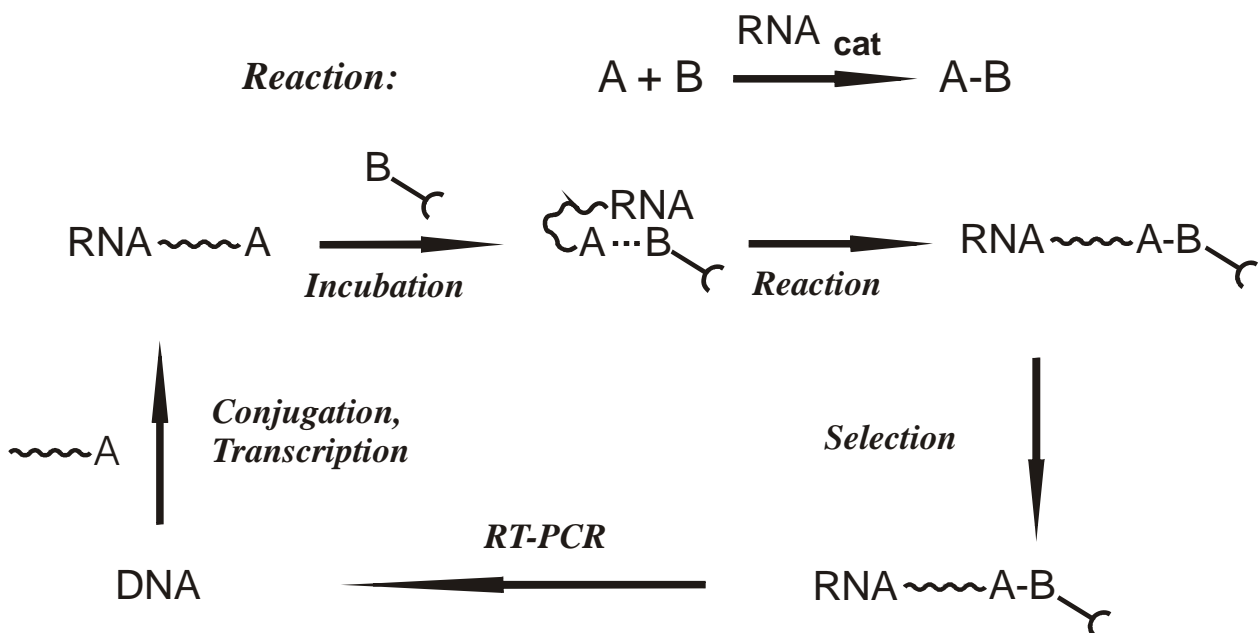
# Problem:

Suitable only for modifying reactions of RNA.

## Challenge:

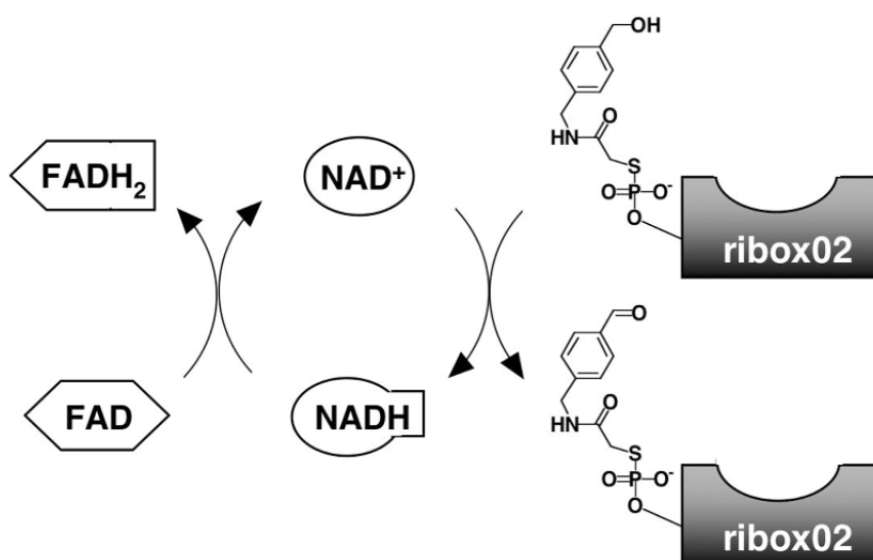
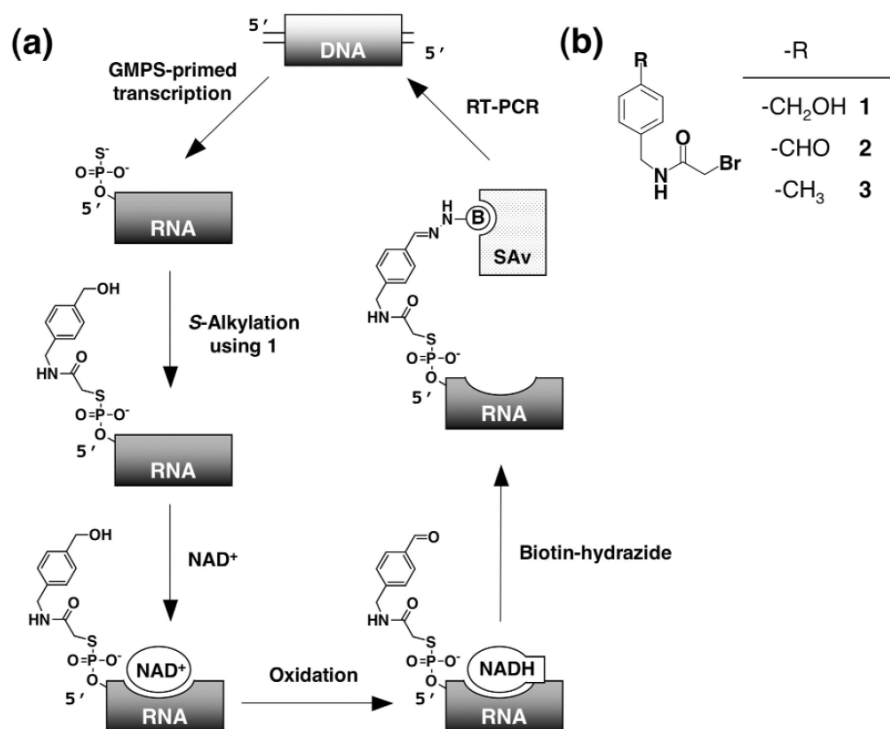


## Direct selection with tethered reactants





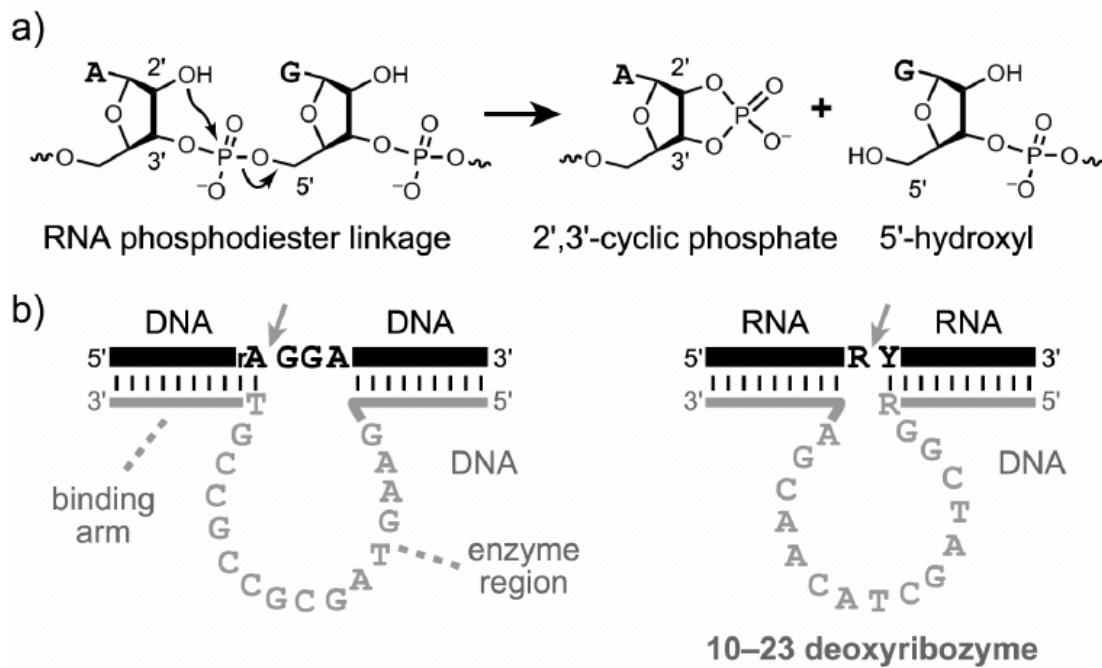
# Example: a redox ribozyme

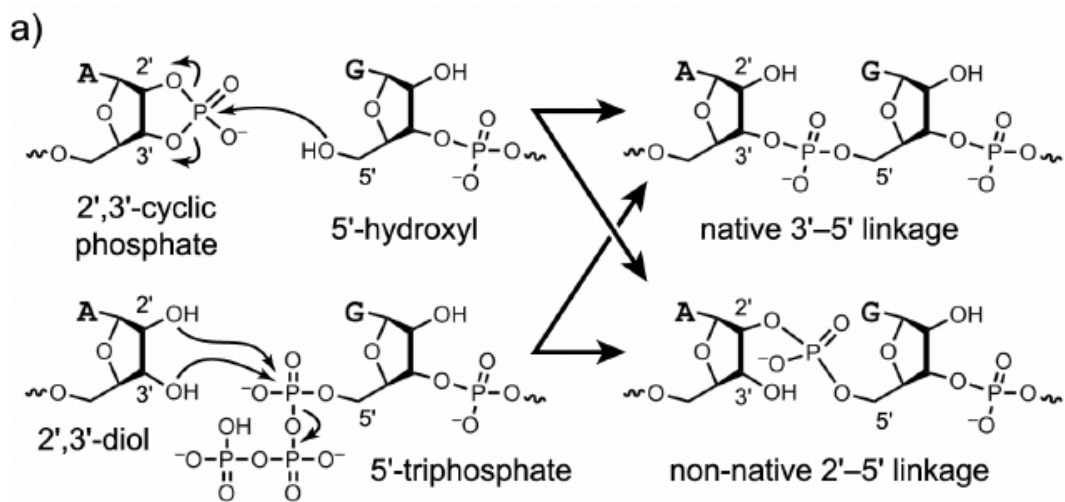


**Fig. 9** A multicomponent redox system involving 1-ribox02, NAD<sup>+</sup>/NADH, and FAD/FADH<sub>2</sub> [31].

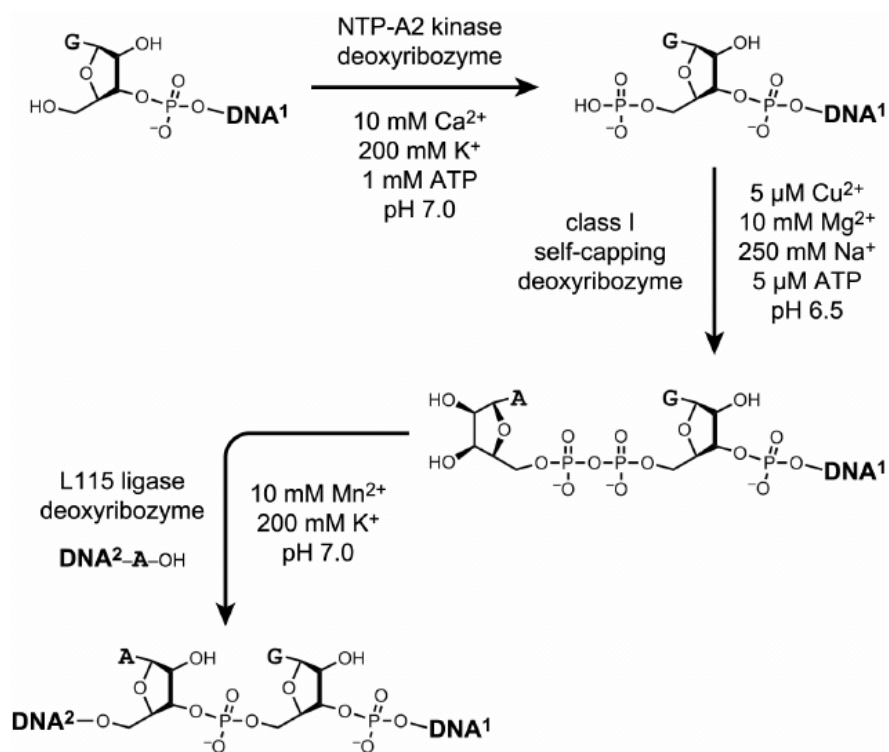
# Deoxyribozymes

- Discovery
- Scope
- Limitations





S.K. Silverman



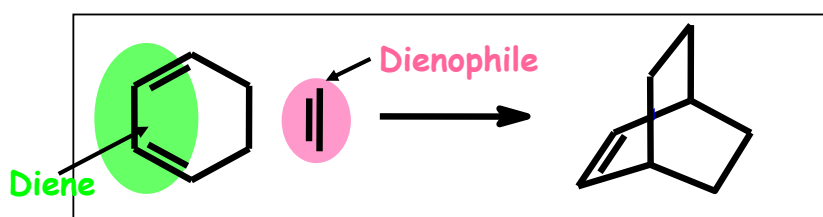
S.K. Silverman

# RNA-catalyzed Diels-Alder reactions

## Rationales

- Exploration of the catalytic bandwidth of RNA
- Understand structural and mechanistic principles
- Compare with protein enzymes, deduce general governing principles

## The Diels-Alder reaction

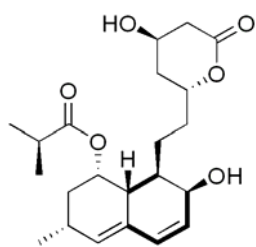


Otto Diels  
(1876-1952)

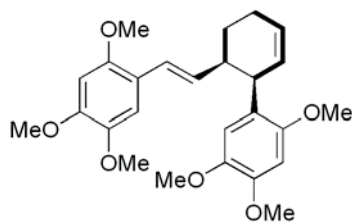
Kurt Alder  
(1902-1958)

# Catalysis of Diels-Alder reactions in Nature

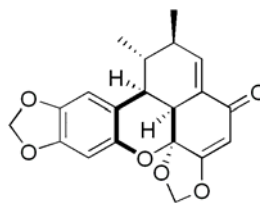
•Secondary metabolites thought to be biosynthesized via Diels-Alder reaction:



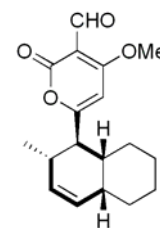
Mevinolin  
(*Aspergillus terreus*)



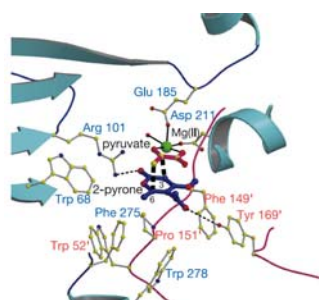
Alflabene  
(*Alpinia flabellata*)



Carpanone  
(*Cinnamomum sp.*)



Solanapyrones A  
(*Alternaria solani*)



Ose et al. *Nature* 2003

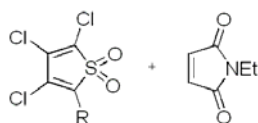
First solved structure of an enzyme which is assumed to catalyze a Diels Alder Reaktion by Ose *et al.*:

•Macrophomate Synthase (MPS)

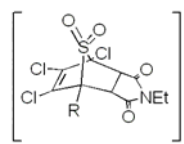
To date, there is no solid proof for the existence of Diels-Alderases in Nature.

## Artificial biopolymeric Diels-Alder catalysts

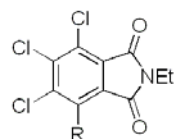
catalytic antibodies  
(Abzymes)



Abzyme  
1E9

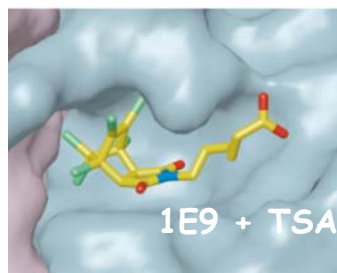


1.- SO<sub>2</sub>  
2. [Ox.]

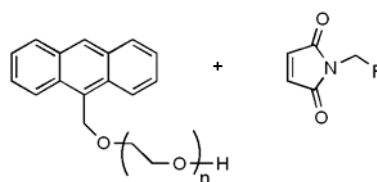


Involved Interactions:

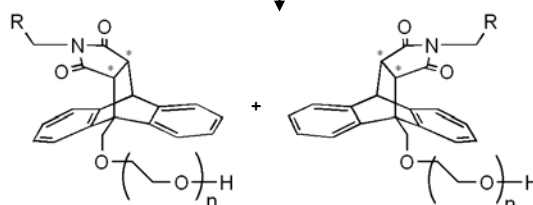
- hydrophobic interactions
- 1 hydrogen bond



catalytic RNA  
(Ribozymes)

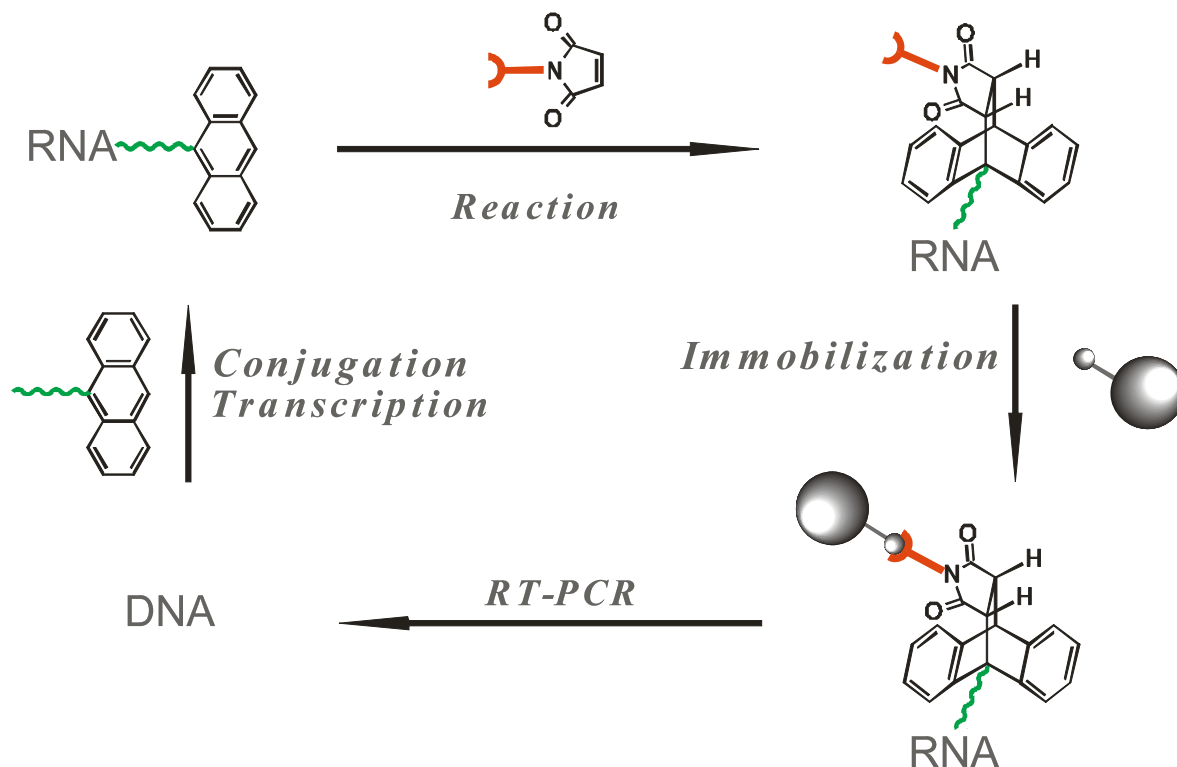


Ribozyme

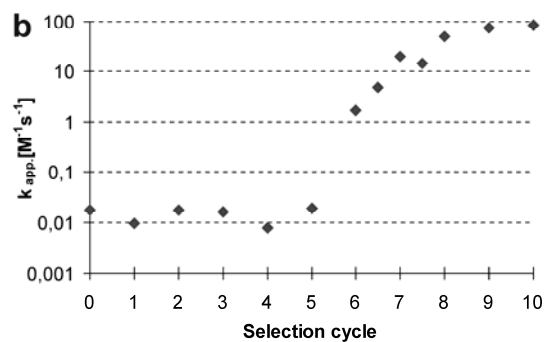
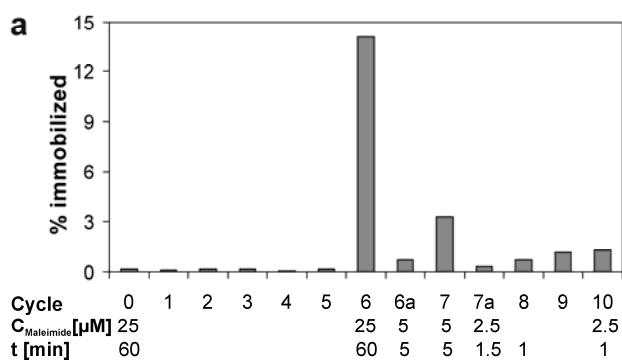


structure and mechanism ?

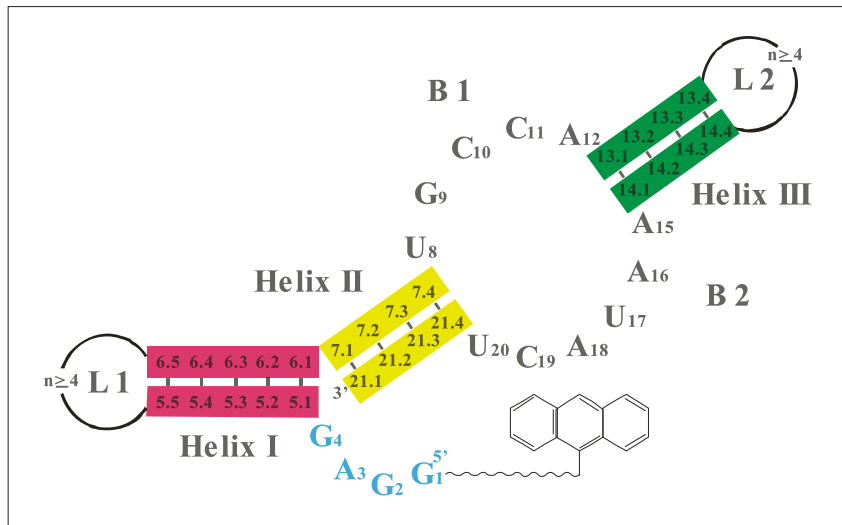
# *In vitro* selection of Diels-Alderase ribozymes



## Progress of the selection

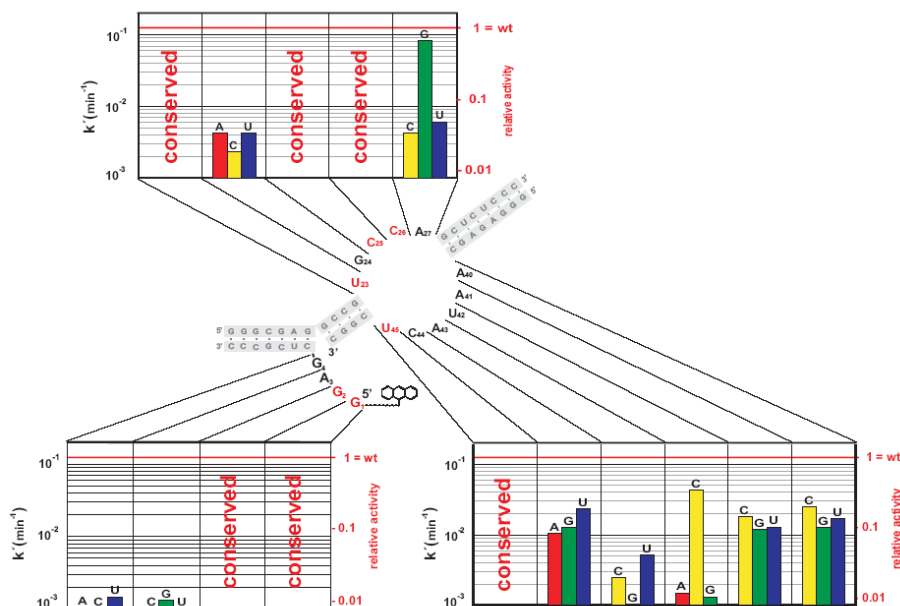


# Catalytic minimal motif



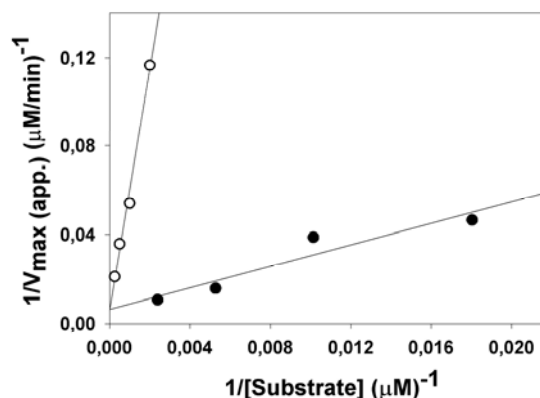
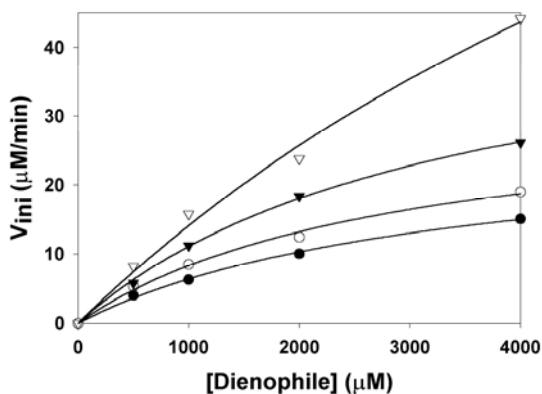
Seelig & Jäschke, *Chem. Biol.* 1999 (6) 167-176

# Probing the secondary and tertiary structure



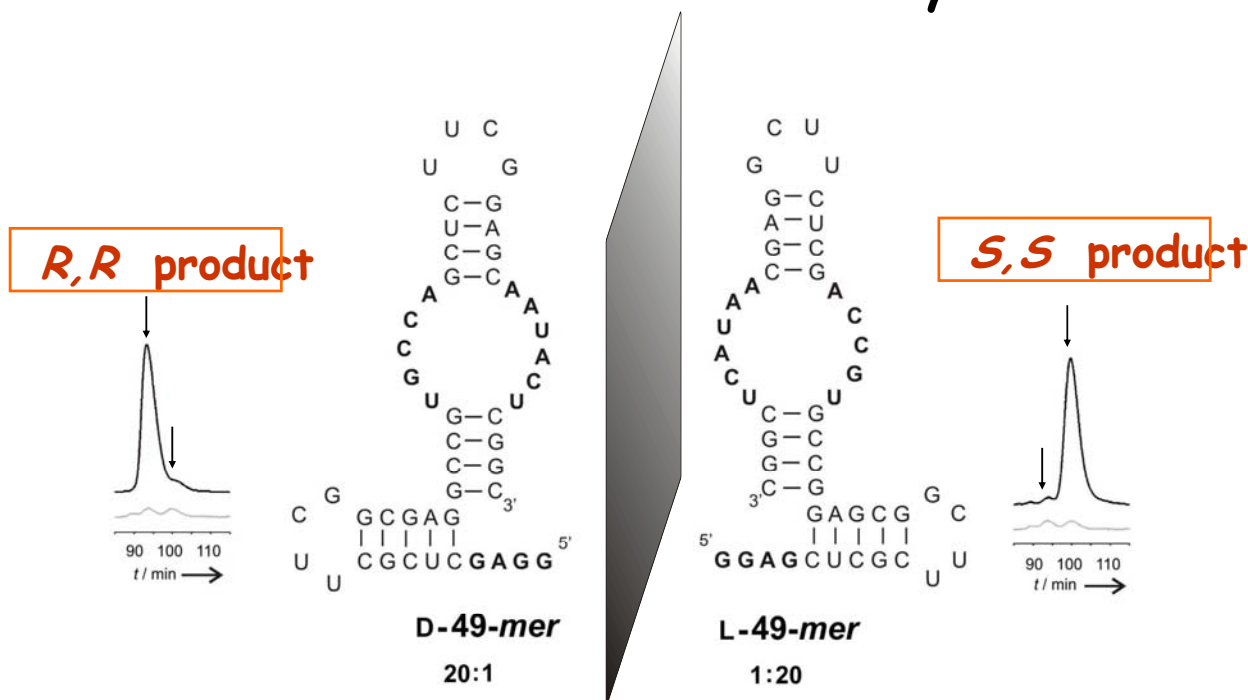
Keiper, Bebenroth, Seelig, Westhof & Jäschke, *Chem. Biol.* 2004 (11) 1217-1227.

# The hallmarks of enzymatic catalysis - saturation behaviour & multiple turnovers



Seelig, B., Keiper, S. Stuhlmann, F. & Jäschke, A. *Angew. Chem.* 2000 (112) 4764-4768.

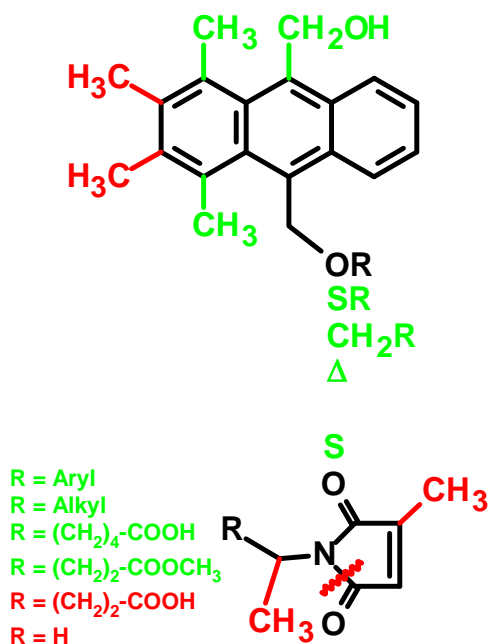
# The hallmarks of enzymatic catalysis - Stereoselectivity



Seelig, B., Keiper, S. Stuhlmann, F. & Jäschke, A. *Angew. Chem.* 2000 (112) 4764-4768.

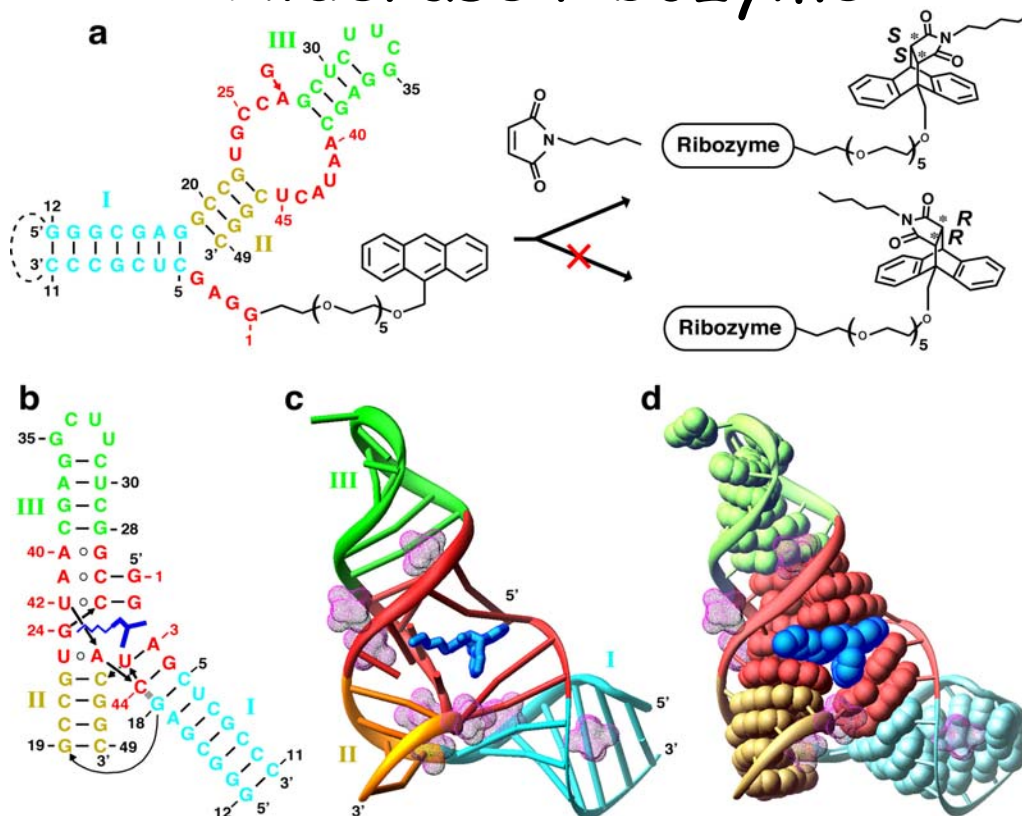


# RNA - substrate interactions

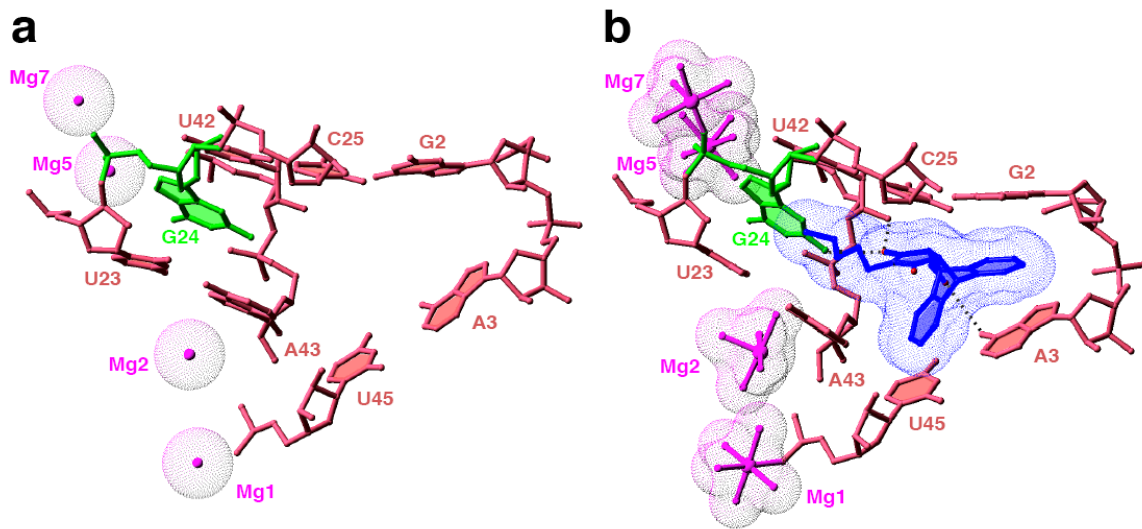


Stuhlmann & Jäschke *J. Am. Chem. Soc.* 2002 (124) 3238-3244.

# The crystal structure of the Diels-Alderase ribozyme

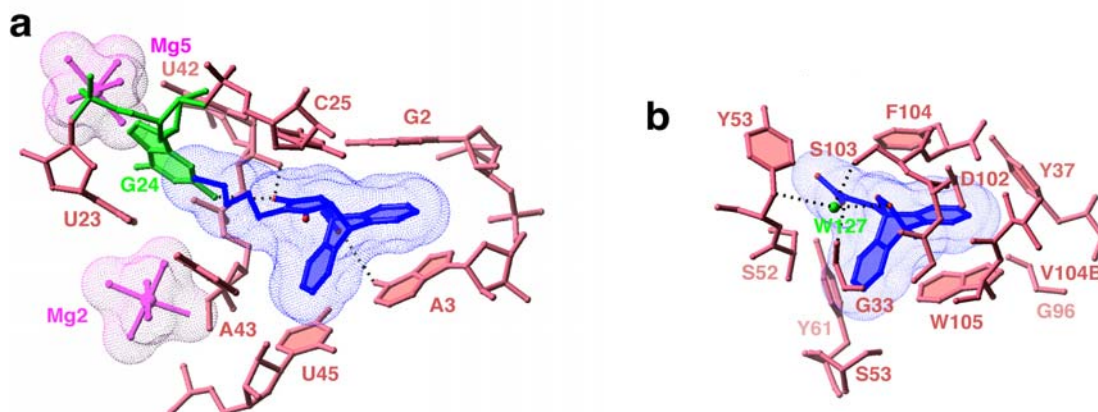


# RNA - product interactions



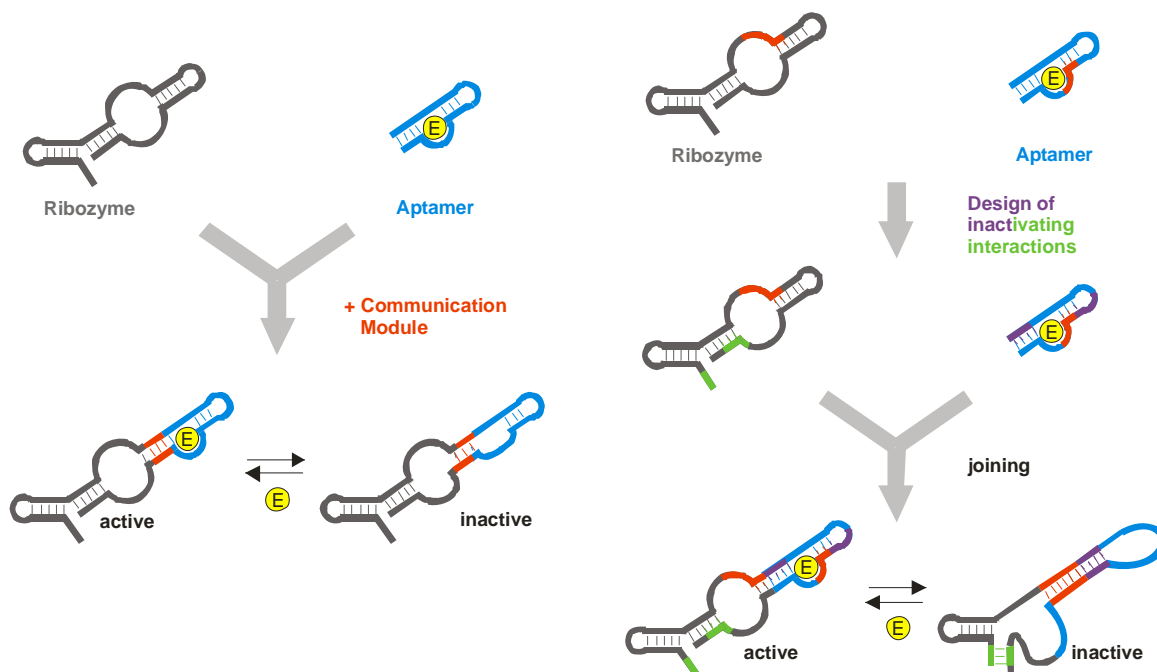
Serganov, A., Keiper, S., Malinina, L., Tereschko, V., Skripkin, E., Höbartner, C., Polonskaia, A., Phan, A. T., Wombacher, R., Micura, R., Dauter, Z., Jäschke, A., Patel, D. J.: *Nature Struct. Mol. Biol.* 2005 (12) 218-224.

## Comparison with Protein Diels-Alderase



Hugot *et al.* *PNAS* 2002 (99) 9674-9678.

# Can we use this structural information for reaction control?

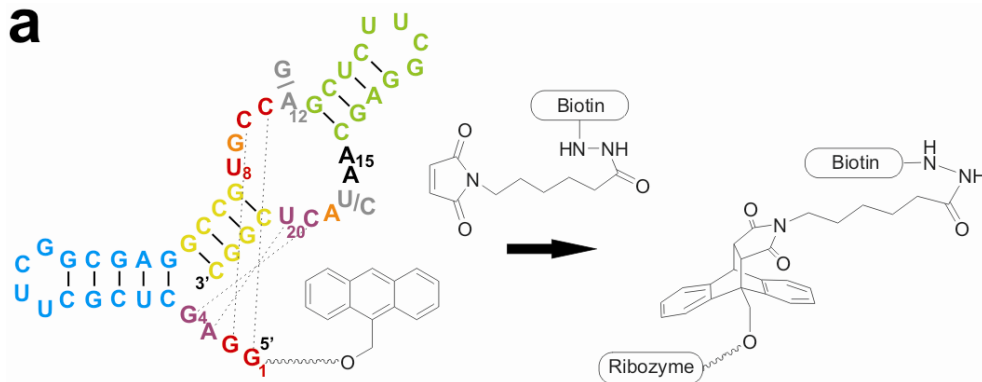


Helm, M., Petermeier, M., Ge, B., Fiammengo, R., Jäschke, A.: *J. Am. Chem. Soc.* **2005** (127) 10492-93.

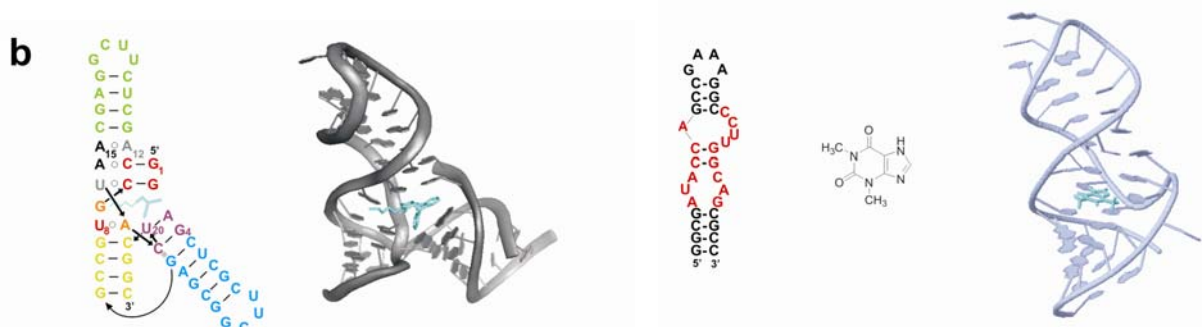
Amontov, S., Jäschke, A.: *Nucleic Acids Res.* **2006** (34) 5032-38; Petermeier, M., Jäschke, A. *Org. Biomol. Chem.* **2009** (7) 288-292.

## Allosteric Diels-Alderase ribozymes

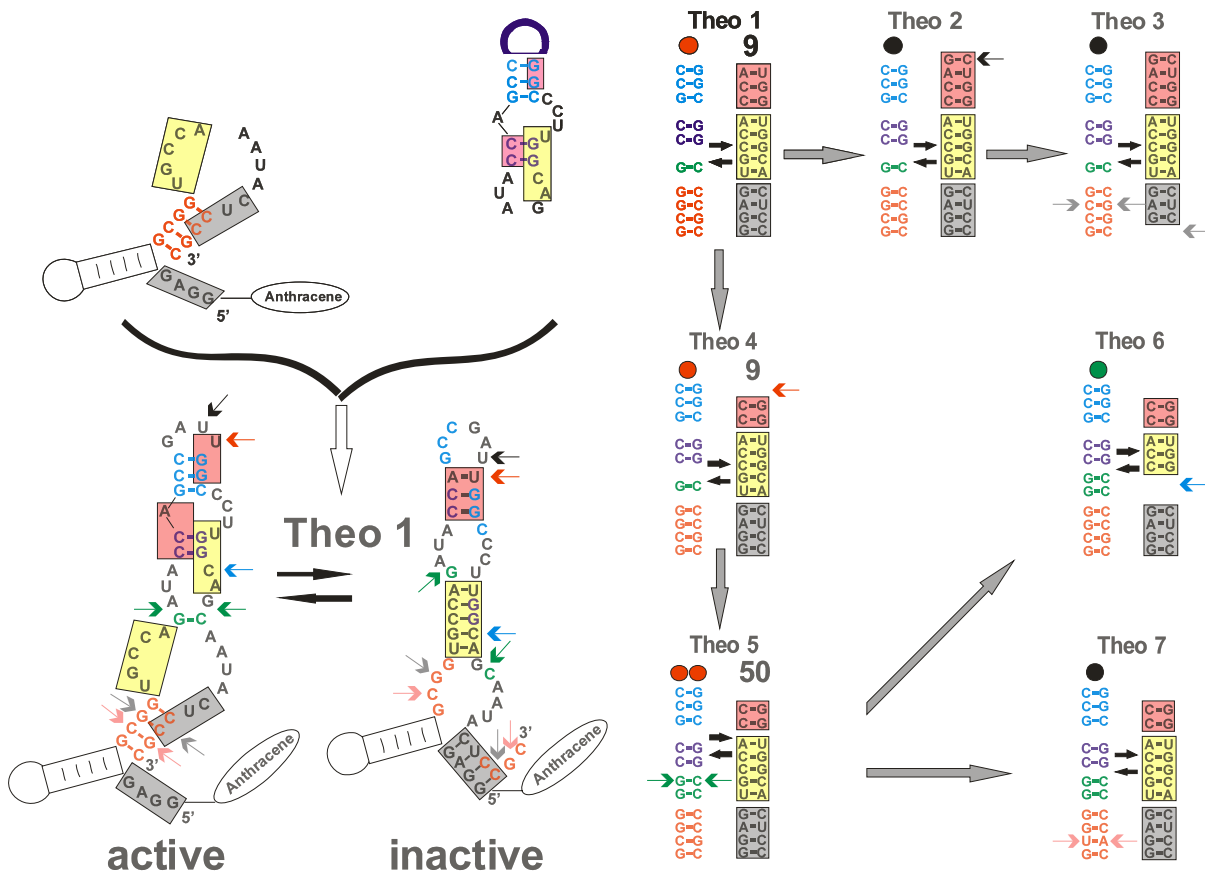
**a**



**b**



Theophylline aptamer



## Theophylline-dependent Diels-Alderase ribozyme

